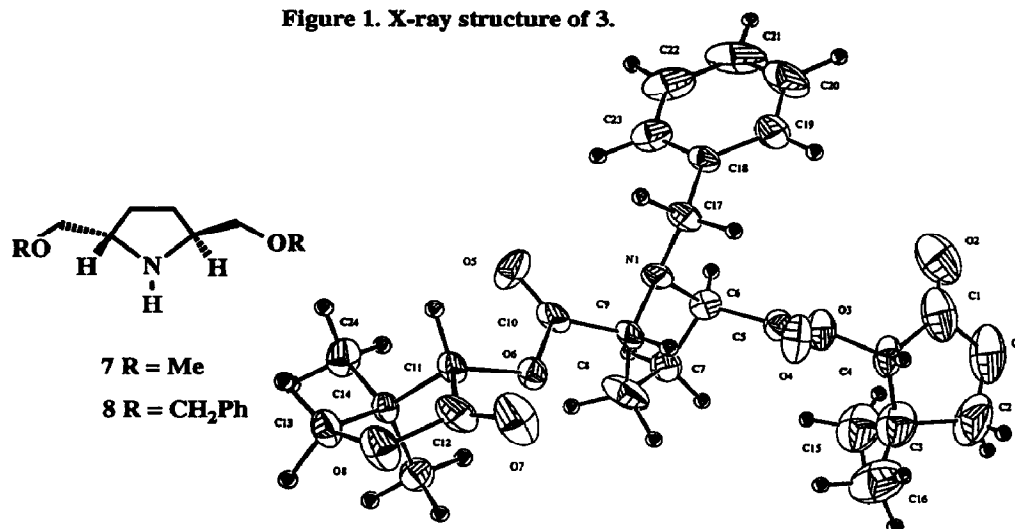


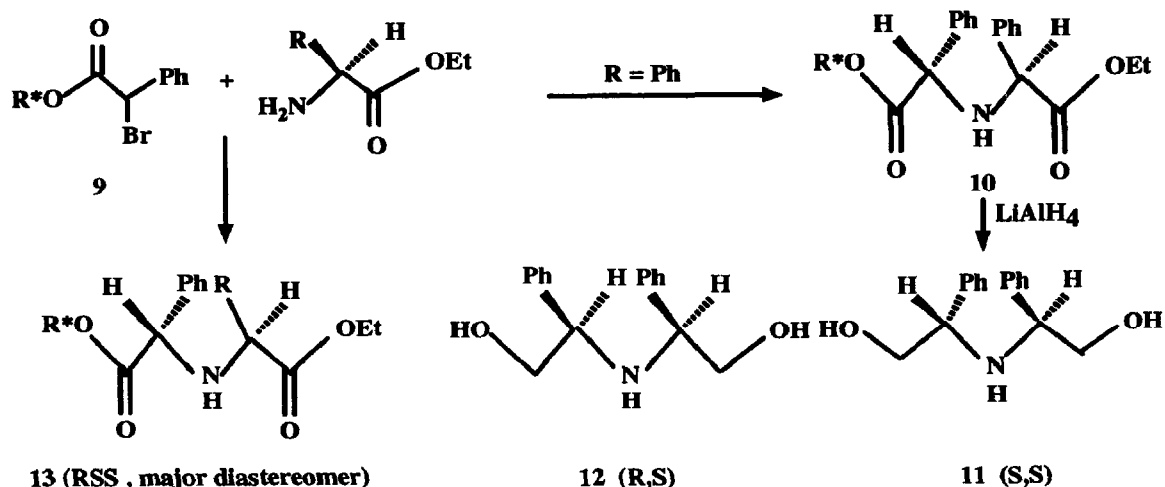
Optically active pyrrolidine-trans-2,5-dicarboxylic acids **6** were first prepared by Katsuki via resolution.<sup>5</sup> This procedure was improved by Ghosez.<sup>6</sup> Yamamoto also used a resolution step to prepare **7**.<sup>7</sup> Synthesis of optically active **7** and **8** have recently be reported starting with (*S*)-*O*-benzylglycidol (8 steps)<sup>8</sup> and D-mannitol (8 steps);<sup>9</sup> **6** has recently been prepared from *N*-BOC pyroglutamate ethyl ester.<sup>10</sup> The present synthesis offers the advantage of conciseness and high selectivity for one of the enantiomers in the pyrrolidine ring forming reactions.

Derivatives of **5** such as **6** and **7** are of considerable interest as  $C_2$ -symmetric chiral auxiliaries<sup>11</sup> since their carboxamide enolates show remarkable stereochemical control in alkylations,<sup>5,12a-c</sup> acylations,<sup>12d</sup> aldol<sup>12e</sup> and Wittig rearrangement reactions.<sup>12f</sup> These auxiliaries have also been used to control the stereochemical outcome of Diels-Alder cycloadditions<sup>12g</sup> and halolactonizations.<sup>12h</sup>

Figure 1. X-ray structure of **3**.



Non-cyclic  $C_2$ -symmetric  $\beta,\beta'$ -dihydroxyamines can also be produced via this methodology. Thus, reaction of **9**, prepared from racemic  $\alpha$ -bromophenylacetic acid with (*S*)-phenylglycine ethyl ester afforded the diester **10** (RSS:RRS ratio=18:1) in 80% isolated yield.<sup>13</sup> Reduction with  $LiAlH_4$  gave the (*S,S*)-amino diol **11**,  $[\alpha]_D^{22} +133.5^\circ$  (c 1.4,  $CH_2Cl_2$ ).<sup>14</sup> Similar treatment of **9** with (*R*)-phenylglycine ethyl ester resulted in 70% of the (RSR) diastereomer of **10**; reduction gave the meso diol **12**  $[\alpha]_D^{22} 0^\circ$  (c 0.85,  $CH_2Cl_2$ ).

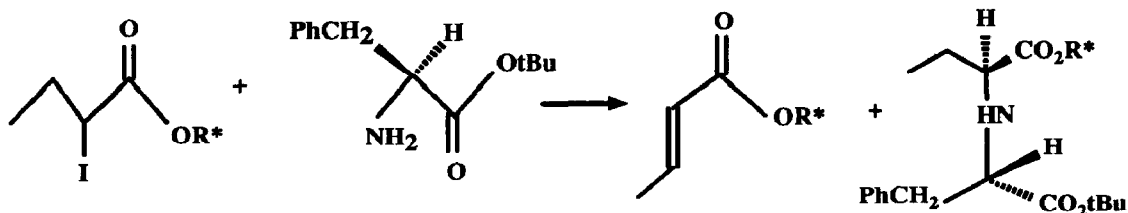


Reaction of **9** with other optically active  $\alpha$ -amino esters gave a series of amino diesters **13** in similar and even higher diastereomer ratios. These results are summarized below.

<u>Amino ester</u>	<u>Isolated Yield (%)</u>	<u>Diastereomer Ratio (RSS:RRS)</u>
(S)-Glycine ethyl ester	70	12:1
(S)-Phenylalanine methyl ester	60	9:1
(S)- Phenylalanine t-butyl ester	77	27:1
(S)-Proline methyl ester	74	10:1
(S)- Proline t-butyl ester	90	8:1

The reduction products derived from **13** no longer have  $C_2$  symmetry. Nevertheless, they show that a variety of optically active  $\beta,\beta'$ -dihydroxyamines can be prepared by this methodology. Hydrolysis of **13** leads to optically active compounds consisting of two  $\alpha$ -amino acids which share the one nitrogen atom present in the molecule. Such compounds are structurally related to the octopines and nopalines which are implicated in crown gall tumors in higher plants.<sup>15</sup> Other related structures have been identified as inhibitors of angiotensin-converting enzyme as well as thermolysin<sup>16</sup>; they are also components of peptide antibiotics.<sup>17</sup>

Initial experiments indicate that displacement of bromide or iodide ion by  $\alpha$ -amino esters from non-aromatic analogs of **9** is accompanied by significant amounts of elimination product. However, the diastereomer enrichment is still in the >15:1 range.



### Acknowledgments

We would like to thank C. Bensimmon for providing the X-ray crystallographic data and the Natural Science and Engineering Research Council of Canada for the financial support of this research

### References and Notes

- φ - NSERC (Canada) PGS Fellow 1992-1994
- Durst, T.; Koh, K., R.N., Ben. *Tetrahedron Lett.*, **1993**, *34*, 4476.
  - All new products described in this article were isolated by chromatography on silica gel or recrystallization, and show satisfactory analytical data (IR, NMR, <sup>13</sup>C and MS). All the diastereomeric ratios of the products were determined by examination of the crude reaction mixtures in CDCl<sub>3</sub> using <sup>1</sup>H NMR (300 MHz or 500 MHz)
  - 3** has the following physical data: Mp 147-148°C. [α]<sub>D</sub><sup>22</sup> -61.7° (c 1.3, CH<sub>2</sub>Cl<sub>2</sub>). IR(CH<sub>2</sub>Cl<sub>2</sub>): 1751, 1796 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 1.10(s, 6H), 1.20(s, 6H), 1.90-2.10(m, 2H), 2.30-2.50(m, 2H), 3.83(d, 1H, J= 13.10 Hz), 3.90-3.97(m, 2H), 4.03(ABq, 4H, J= 9.04 Hz), 4.08(d, 1H, J= 13.10 Hz), 5.37(s, 2H), 7.11-7.50(m, 5H). <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>): δ 20.08(CH<sub>3</sub>), 23.06(CH<sub>3</sub>), 28.70(CH<sub>2</sub>), 40.10(C), 53.67(CH<sub>2</sub>), 63.01(CH), 75.10(CH), 76.14(CH<sub>2</sub>), 127.35(CH), 128.41(CH), 129.10(CH), 137.85(C), 171.92(C), 172.85(C). EIMS *m/z*: 316(77), 91(100). CIMS *m/z*: 474(100, M+1), 316(77).
  - Physical data: [α]<sub>D</sub><sup>22</sup> -36.2° (c 0.43, CH<sub>2</sub>Cl<sub>2</sub>). IR(CH<sub>2</sub>Cl<sub>2</sub>): 3373 cm<sup>-1</sup>. <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>): δ 1.60-2.10(m, 4H), 2.84(bs, 2H), 3.49(dd, 2H, J= 3.28, 11.01 Hz), 3.58(dd, 2H, J= 4.56, 11.01 Hz), 3.86(ABq, 2H, J= 13.92 Hz), 7.10-7.50(m, 5H). <sup>13</sup>C NMR (50 MHz, CDCl<sub>3</sub>): δ 27.47(CH<sub>2</sub>), 52.25(CH<sub>2</sub>), 62.28(CH<sub>2</sub>), 62.95(CH), 127.79(CH), 128.80(CH), 129.11(CH), 139.62(C). EIMS *m/z*: 190(47), 91(100). CIMS *m/z*: 222(100, M+1), 204(82), 190(95).
  - Kawanami, Y.; Ito, Y.; Kitagawa, T.; Yaniguchi, Y.; Katsuki, T.; Yamaguchi, M. *Tetrahedron Lett.*, **1984**, *25*, 857.
  - Chen, L.Y.; Ghosez, L. *Tetrahedron Lett.*, **1990**, *31*, 4467.
  - Yamamoto, Y.; Hoshino, J.; Fujimoto, Y.; Ohmoto, J.; Sawada, S. *Synthesis*, **1993**, *3*, 298.
  - Takano, S.; Moriya, M. Iwabuchi, Y.; Ogasawara, K. *Tetrahedron Lett.*, **1989**, *30*, 3805.
  - Marzi, M.; Misiti, D. *Tetrahedron Lett.*, **1989**, *30*, 6075.
  - Ezpuerra, J.; Rubio, A.; Pedregal, C.; Sanz, G.; Rodriguez, J.H.; Garcia Ruano, J.L. *Tetrahedron Letters*, **1993**, *34*, 4989.
  - Whitesell, K.W. *Chem. Rev.*, **1989**, *89*, 1581.
  - a) Ikegami, S.; Hayama, T.; Katsuki, T.; Yamaguchi, M. *Tetrahedron Lett.*, **1986**, *27*, 3403. b) Enomoto, M.; Ito, Y.; Katsuki, T.; Yamaguchi, M. *Tetrahedron Lett.*, **1985**, *26*, 1343. c) Hanamoto, T.; Katsuki, T.; Yamaguchi, M. *Tet. Lett.*, **1986**, *27*, 2463. d) Ito, Y.; Katsuki, T.; Yamaguchi, M. *Tetrahedron Lett.*, **1984**, *25*, 6015. e) Katsuki, T.; Yamaguchi, M. *Tetrahedron Lett.*, **1985**, *26*, 5807. f) Uchikawa, M.; Hanamoto, T.; Katsuki, T.; Yamaguchi, M. *Tetrahedron Lett.*, **1986**, *27*, 4577. g) Gouverneur, V.; Ghosez, L. *Tetrahedron Lett.*, **1991**, *32*, 5349. h) Fuji, K.; Nodo, M.; Naniwa, Y.; Kawabata, T. *Tetrahedron Lett.*, **1990**, *31*, 3175.
  - 10** has the following physical data: [α]<sub>D</sub><sup>22</sup> +1.1° (c 10.4, CH<sub>2</sub>Cl<sub>2</sub>). IR (CH<sub>2</sub>Cl<sub>2</sub>): 1072, 1168, 1691, 1745, 1797, 2912, 3347 cm<sup>-1</sup>. <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>): δ 0.60 (s, 3H), 1.00 (s, 3H), 1.21 (t, 3H, J= 8.70 Hz), 3.95 (q, 2H, J= 8.70 Hz), 4.05 (m, 2H), 4.12 (s, 1H), 4.45 (s, 1H), 5.26 (s, 1H), 7.42 (m, 1H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 13.84, 18.86, 22.55, 40.33, 61.21, 61.90, 62.05, 75.14, 75.83, 75.90, 127.71, 128.00, 128.26, 128.53, 128.73, 137.03, 137.09, 171.30, 171.90. CIMS *m/z*: 426 (100, M+1), 412 (12), 352 (55), 247 (30), 131 (22), 105 (24).
  - 11** has the following physical data: [α]<sub>D</sub><sup>22</sup> +133.5° (c 1.4, CH<sub>2</sub>Cl<sub>2</sub>). IR (CH<sub>2</sub>Cl<sub>2</sub>): 1041, 1493, 2354, 2930, 3600 cm<sup>-1</sup>. <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>): δ 2.80 (bs, 3H), 3.50-3.80 (m, 6H), 7.10-7.40 (m, 10H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>): δ 60.73, 66.66, 127.14, 127.49, 128.39, 139.15. CIMS *m/z*: 258 (20, M+1), 240 (12), 226 (30), 102 (25).
  - Drummond, M. *Nature*, **1979**, *281*, 343.
  - Miyazawa, T.; Hiramatsu, S.; Yasuhiro, T.; Yamada, T.; Kuwata, S. *Bull. Chem. Soc. Jpn.*, **1985**, *58*, 1976.
  - Miyazawa, T. *Bull. Chem. Soc. Jpn.*, **1980**, *53*, 2555.